# Reaction Kinetics of Oxyphosphoranes with Model Compounds of Nucleosides

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Received September 27, 1996

Ester exchange reaction kinetics of oxyphosphoranes with several kinds of alcohols, the model compounds for nucleosides, has been investigated. Comparison of the reaction rates of oxyphosphorane (1) with alcohols indicated that the ester exchange rates of diols were much faster than that of monoalcohols. These results might provide a clue for the intrinsic difference between ribonucleotides and 2'-deoxyribonucleotides. © 1997 Academic Press

#### INTRODUCTION

Ribonucleotides and 2'-deoxyribonucleotides showed great differences in chemical and biochemical properties, mainly because of the existence of the 2'-OH group in ribonucleotides. In the literature, phosphoranes had been proposed as the possible intermediates for the displacement reaction of nucleotides with water, alcohols, and other nucleophiles (1–7). In order to clarify the intrinsic different properties of ribonucleotides and 2'-deoxyribonucleotides, some simple alcohols were chosen as the model compounds for nucleosides and reacted with oxyphosphoranes. For example, ethylene glycol was chosen as the 2',3'-diOH moiety of ribonucleosides, 1,3-propanediol as the 3',5'-diOH moiety, 1,2,3-propanetriol as the 2',3',5'-triOH moiety, and ethanol, n-propanol as the 5'-OH moiety. Their structure and reactivity relationship could provide some understanding of the chemistry in life chemistry (7).

#### RESULTS AND DISCUSSION

Reaction Kinetics of Oxyphosphorane (1) with Diols and Triol

Oxyphosphorane (1) (8) dissolved in pyridine was reacted at room temperature with one molar equivalent of ethylene glycol to give a spirooxyphosphorane (2) in an almost quantitative yield (Scheme 1). The reaction proceeded so rapidly that 1 was consumed completely within 1 h. Ramirez *et al.* (9) obtained this spirooxyphosphorane from a three-step procedure in less than 50% yield.

Under the similar conditions, oxyphosphorane (1) was reacted with 1,3-propanediol within 5 h to afford a spirooxyphosphorane attached by a six-membered ring (1,3,2-dioxaphosphorinane ring) (3) in about 95% yield. The mass spectrum of 3,

SCHEME 1. Reaction of oxyphosphorane (1) with ethylene glycol, 1,3-propanediol, ethanol and n-propanol (pyridine, 22°C).

which displayed a molecular ion peak at m/z 346, and its <sup>31</sup>P NMR shift at -49.05 ppm, confirmed the structure.

The disappearance rate constants under different temperatuare for oxyphosphorane (1) as it was reacted with ethylene glycol and 1,3-propanediol respectively were calculated by means of <sup>31</sup>P NMR spectra, and summarized in Table 1. At 0°C, the 1,2-diol reacted with 1 about 7.2 times faster than 1,3-diol. As the temperature raised to 22°C and 38°C, respectively, their absolute rate increased; however, the relative reactivities of 1,2-diol to 1,3-diol had decreased to 4.1 and 2.9, respectively. The difference of the activation energy Ea for the reaction of oxyphosphorane 1 with 1,2-diol and 1,3-diol was about 4.0.

When equal amounts of ethylene glycol and 1,3-propanediol were competed for the substrate 1, after equilibrium, the product 2 was the major one in 92%, while 3 was the minor one in 8%. The ratio of 2 to 3 was about 12:1 (Scheme 2).

When 1,2,3-propanetriol was employed to proceed the ester exchange reaction,

TABLE 1							
Thermodynamic Parameters for the Reaction of Oxyphosphorane (1) with Ethylene Glycol and							
1,3-Propanediol <sup><math>a</math></sup>							

Alcohol	$k \times 10^4$						
	0°C	22°C	38°C	Ea	$\Delta H$	$\Delta S$	$\Delta G$
1,2-diol	9.78	31.2	65.5	8.46	7.87	-43.3	20.6
1,3-diol	1.35	7.55	22.6	12.5	11.9	-32.5	21.5

<sup>&</sup>lt;sup>a</sup> Unit of k, Ea ( $\Delta$ H,  $\Delta$ G), and  $\Delta$ S is  $M^{-1}.S^{-1}$ , kcal.mol<sup>-1</sup> and cal.deg<sup>-1</sup>.mol<sup>-1</sup>, respectively.

the <sup>31</sup>P NMR signals at -26.68 and -27.24 ppm for spirooxyphosphoranes with two five-membered rings (4) appeared first within 1 min. The signals at -49.33 and -50.04 ppm for spirooxyphosphoranes attached by six-membered rings (5) were emerged later. As the reaction was going on, the amounts of 4 increased quickly. Meanwhile, 5 increased very slowly to reach the maximum within 25 min, then decreased gradually, and finally converted into 4. The results were shown in Fig. 1, which were much similar than those when 1 was reacted with ribonucleosides (Scheme 2) (10). The structures of compounds 4 were determined by <sup>31</sup>P, <sup>13</sup>C NMR spectral, and HRMS data. The <sup>1</sup>H NMR signals of 4 were serious overlapping, whereas the <sup>13</sup>C NMR spectrum has fairly good pattern to assign for the diastereomers 4a and 4b (Fig. 2).

### Reaction Kinetics of Oxyphosphorane (1) with Monoalcohols

The nucleophilic substitution of oxyphosphorane (1) with a monohydroxyl compound such as ethanol proceeded much slower, compared to the 1,2-diol and 1,3-diol (Fig. 3). For example, at 22°C, within 1 h ethylene glycol reacted with 1 completely, while only about 4% of ethanol was reacted. It took 30 h to consume 60% of 1 when reacted with ethanol, and there were three products with the  $^{31}P$  NMR signals at -49.47, -50.37, and -51.13 ppm. Their mass spectrum had three molecular ion peaks at m/z 348, 362, and 376, for the mono-, di-, and tri-ester exchanged products 8, 9, and 10, respectively (Scheme 1). The structure of 10 was confirmed by an authentic sample. The ratio of 8 to 9 to 10 was 10:5:1. When the exchanged alcohol was a larger molecule n-propanol, the substitution rate was expected to be slower than ethanol. Indeed, 30 h later, about 50% of the reactant 1 still remained. The  $^{31}P$  NMR spectrum of the reaction mixture revealed three singlets at -49.50, -50.30, and -51.11 ppm in a ratio of 15:7:1, which were due to the mono-, di-, and tri-ester exchanged products 11, 12, and 13 with molecular ion peaks at m/z 362, 390, and 418, respectively, observed in the EIMS.

## Reaction Kinetics of Other Oxyphosphoranes with 1,2-Diol

It is obvious that the 1,2-diol is an unique reactant toward oxyphosphorane (1). Could it only be due to the formation of an extra five-membered ring? However,

SCHEME 2. Reaction regioselectivity of oxyphosphorane (1) toward ribonucleosides and their model compounds.

when oxyphosphorane (14) (11) with a saturated five-membered ring and electronwithdrawing groups (p-nitrophenyl) was reacted with ethylene glycol (Scheme 3), the reaction rate  $(k = 4.25 \times 10^{-6} \text{ m}^{-1}.\text{s}^{-1})$  was 734 times slower than that of oxyphosphorane (1) with an unsaturated five-membered ring. Whereas the reaction rate  $(4.91 \times 10^{-4} \,\mathrm{M}^{-1}.\mathrm{s}^{-1})$  of oxyphosphorane (16) with an unsaturated fivemembered ring attached by electron-donating groups (methyl) was only six times slower than that of oxyphosphorane (1). Hence, it is the double bonds, not the electron-withdrawing or electron-donating groups, of the five-membered rings in oxyphosphoranes, that play the key role during the ester exchange reaction of oxyphosphoranes. Since ribonucleotides have much higher reactivities than 2'deoxyribonucleotides, it might be due to the participation of the 2'-OH in ribonucleotides. According to the literature, hydrolysis of RNA proceeds via cyclic oxyphosphorane transition states (12–15). Therefore from the results described above, it can be deduced that the sugar rings of nucleotides could be considered as the "rigid double bonds." When ribonucleotides (A) are hydrolyzed, the oxyphosphoranes transition state with a five-membered ring (B) are formed, in which the sugar ring